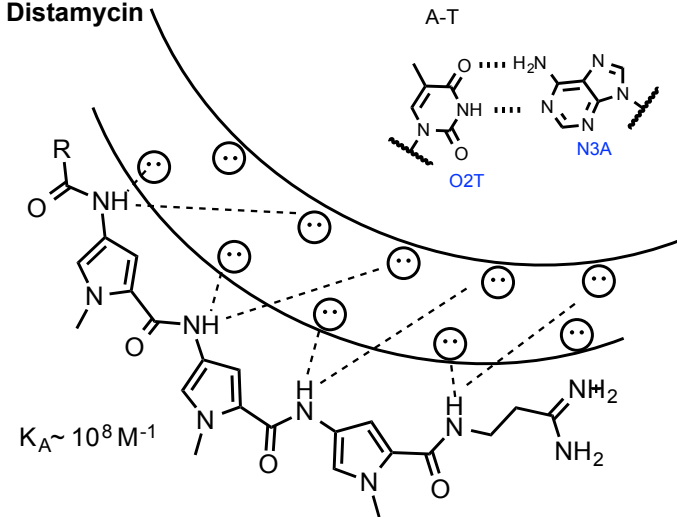


Lecture 3 - Binding of small molecules to DNA

Binding of small molecules to DNA

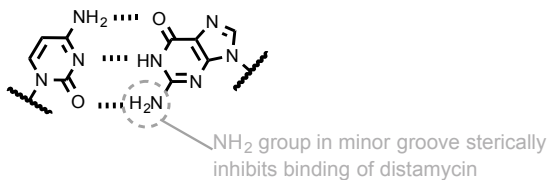
- Electrostatics
- Intercalation
- Groove binding through Van der Waals and hydrogen bonding

Distamycin



- Sequence specific - favors $[A:T]_n$ or $[T:A]_n$, $n=4,5$.
- Binds and displaces the spine of hydration in the minor groove.
- Bifurcated hydrogen bonding of one NH to two lone pairs
- Entropic driving force due to displacement of H_2O .
- Enthalpic driving through Van der Waals and hydrogen bonding.
- Can also have 2:1 binding mode (2 distamycin/site)
- No major change in DNA upon binding

Why not G:C?

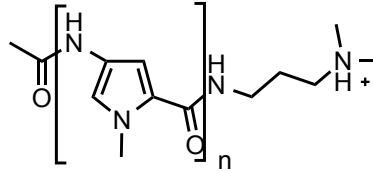


Distamycin-Increasing selectivity

- Selectivity is not great i.e. $(1/2)^5$ - one in every 32 sites is an AT rich run of 5 bases

How do you increase selectivity?

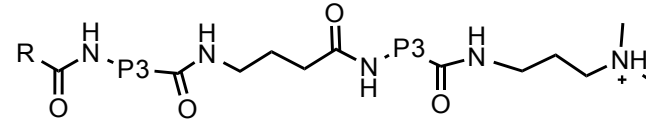
- Simplest solution is to increase the length of polypyrrole



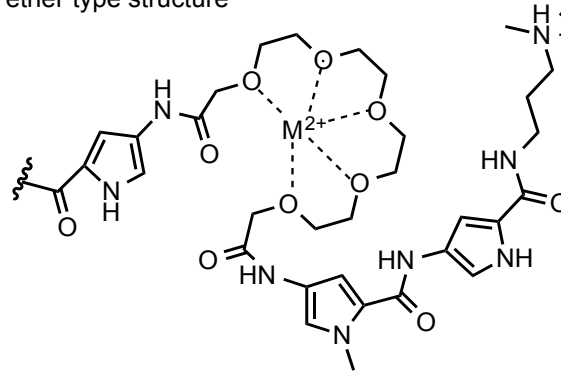
Distamycin = P3 ($n=3$)

This works for $n \leq 5$, but for $n > 5$ the curvature of the polypyrrole backbone doesn't match the curvature of DNA.

- To solve this add in an aminobuteric linker to reorient the pyrrole units

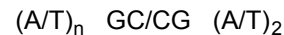


- One can also make a distamycin whose binding is dependent on metal ion binding
- Add in a metal binding sequence i.e. a PEG spacer which will bind to a metal ion to form a crown-ether type structure



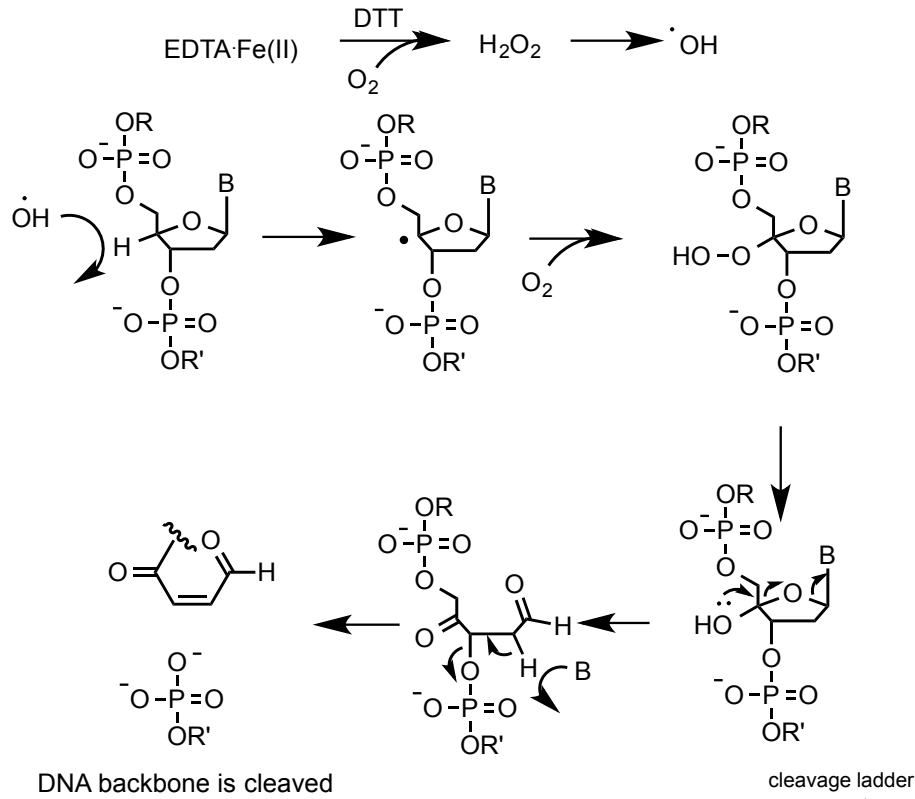
- Sr^{2+} and Ba^{2+} show high binding affinity
- Unfortunately this doesn't work with Ca^{2+}

- Another solution would be to space distamycin with an interchelator

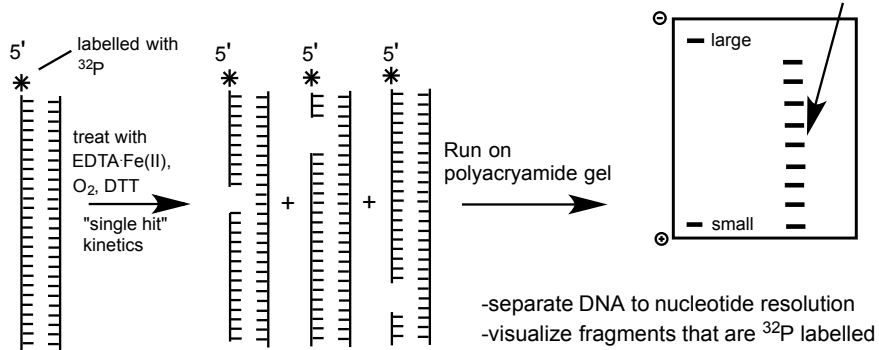


Sequence Specific Binder

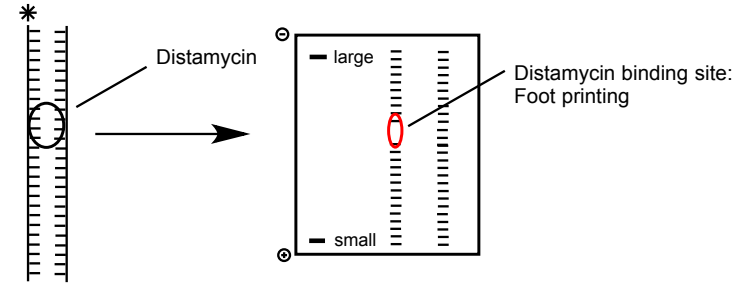
- How do you determine where small molecules bind?
- Take advantage of the following reaction:



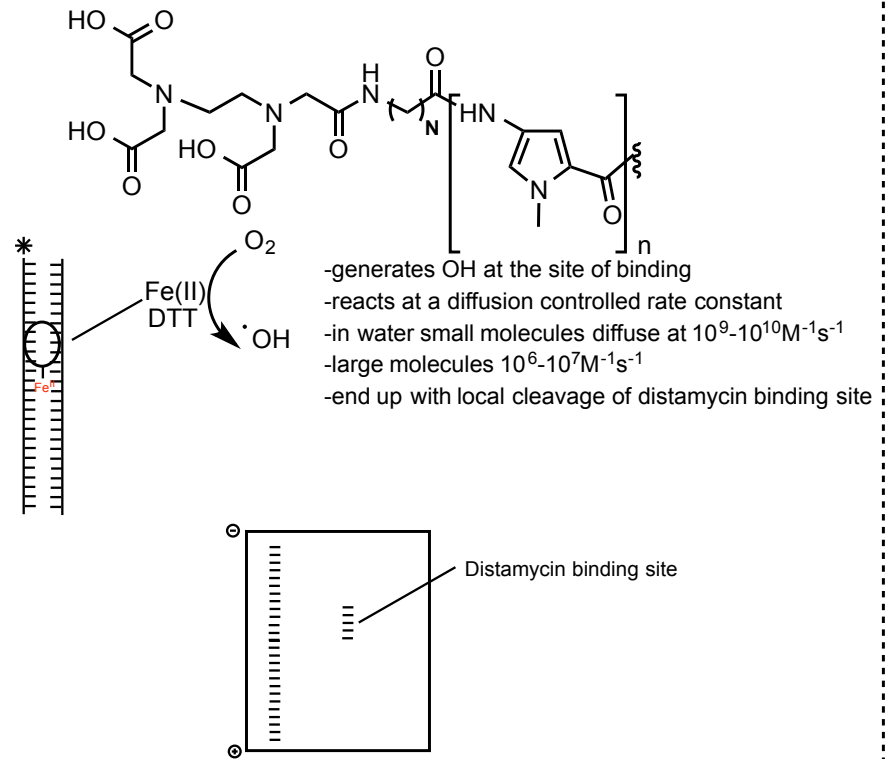
DNA backbone is cleaved



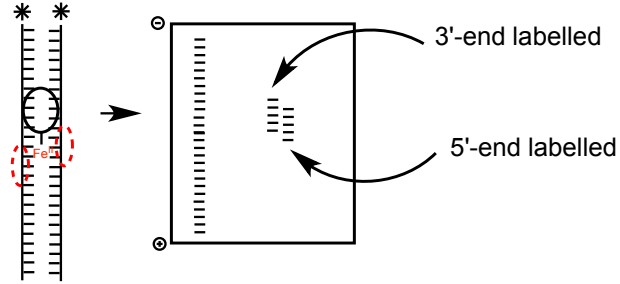
- If you bind a molecule (distamycin) to the DNA, it will provide either steric blockage or quench the cleavage reaction
- The resulting gel will have a "foot-printed" binding site



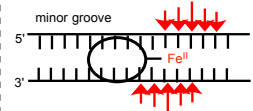
- To increase resolution add an Fe(II) binding motif to generate site specific ·OH adjacent to ligand binding site.



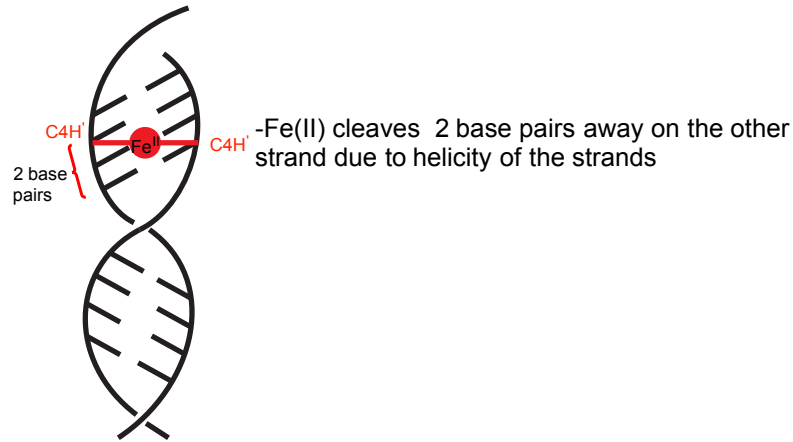
-If you label both strands you can track cleavage on both strands



-observed that cleavage on the 5' strand was shifted in the 3' direction

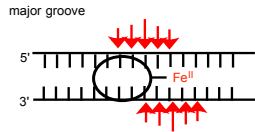


-this is due to the helicity of right-handed DNA

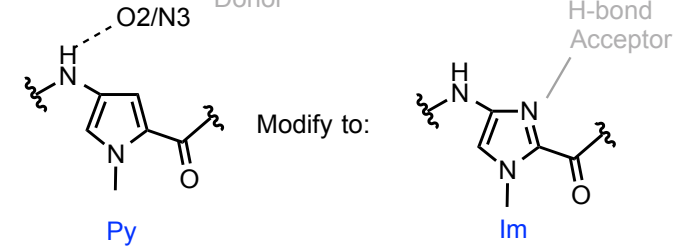
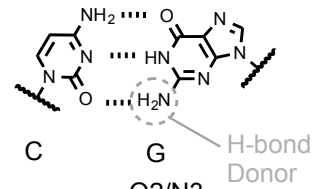


-But helicity in the major groove is different, so Fe(II) cleavage of major groove binders results in a shift in the 5' direction

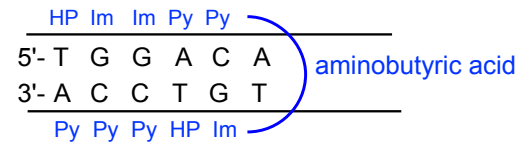
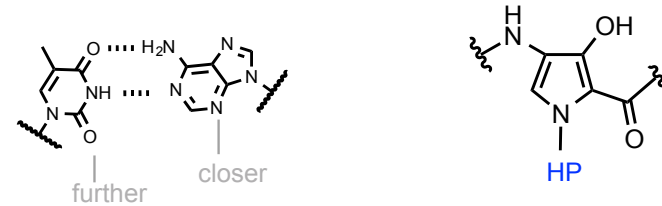
-used to distinguish between major or minor groove binders



-but still limited to AT base pairs



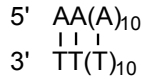
-TA vs. AT



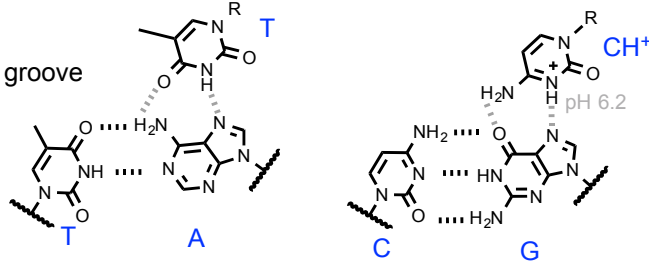
Im/Py GC
 Py/Im CG
 HP/Py TA
 Py/HP AT

Typically bind with $K_D < 10^{-9} M^{-1}$

Binding of longer sequences

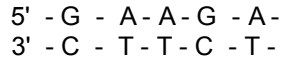
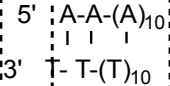
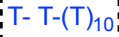


major groove

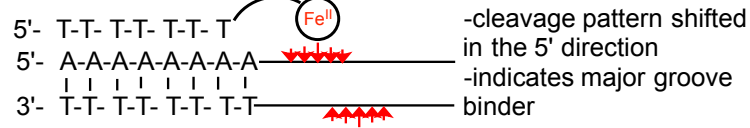


minor groove

- Use Hoogsteen base pairing in the major groove
- Two hydrogen-bonds from protonated CH⁺ (pH 6.2) binding to GC
- Two hydrogen-bonds from T binding to AT

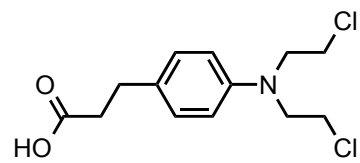


- This only works for $-(\text{Py})_n-$ or $-(\text{Pu})_n-$ sequences (i.e. GATGA does not work).



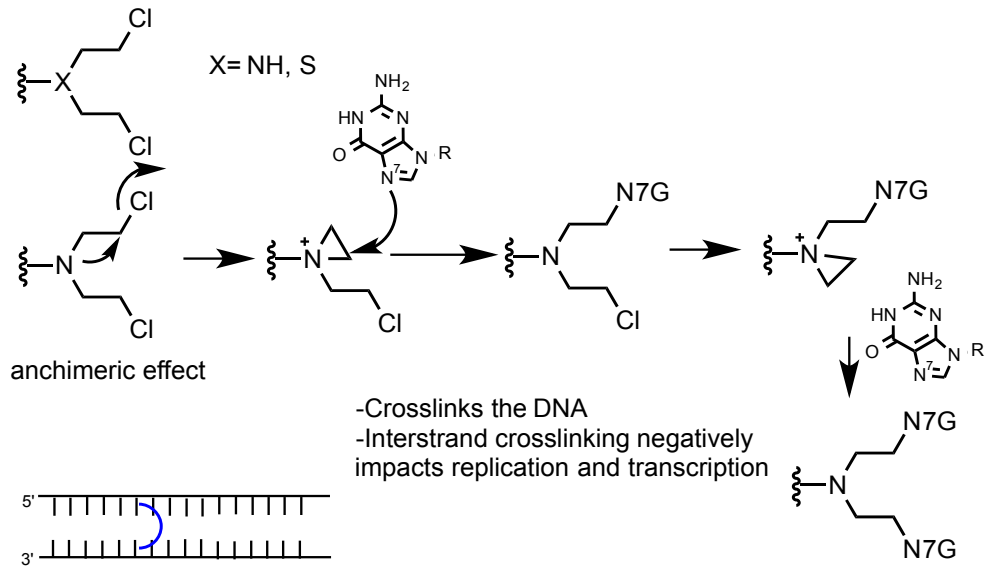
- Hoogsteen is parallel Py/Pu versus antiparallel Py/Pu

Chemical Modification of DNA



- Chlorambucil - anti-cancer drug
- Alkylates the N7 on G

Mustards



G N7 Alkylation leads to tautomerization and mispairing

